



Sgouridis, F., & Ullah, S. (2017). Soil Greenhouse Gas Fluxes, Environmental Controls, and the Partitioning of N₂O Sources in UK Natural and Seminatural Land Use Types. *Journal of Geophysical Research: Biogeosciences*, 122.
<https://doi.org/10.1002/2017JG003783>

Publisher's PDF, also known as Version of record

License (if available):
CC BY

Link to published version (if available):
[10.1002/2017JG003783](https://doi.org/10.1002/2017JG003783)

[Link to publication record in Explore Bristol Research](#)
PDF-document

This is the final published version of the article (version of record). It first appeared online via Wiley at <http://onlinelibrary.wiley.com/doi/10.1002/2017JG003783/abstract> . Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/red/research-policy/pure/user-guides/ebr-terms/>

RESEARCH ARTICLE

10.1002/2017JG003783

Key Points:

- Up to 60% of mean nitrous oxide emissions were attributed to denitrification
- Soil moisture was the key driver regulating the partitioning of N₂O sources
- Methane emissions can be predicted by soil moisture content across ecosystem types

Supporting Information:

- Supporting Information S1

Correspondence to:

F. Sgouridis,
f.sgouridis@bristol.ac.uk

Citation:

Sgouridis, F., & Ullah, S. (2017). Soil greenhouse gas fluxes, environmental controls, and the partitioning of N₂O sources in UK natural and seminatural land use types. *Journal of Geophysical Research: Biogeosciences*, 122. <https://doi.org/10.1002/2017JG003783>

Received 20 JAN 2017

Accepted 14 SEP 2017

Accepted article online 25 SEP 2017

Soil Greenhouse Gas Fluxes, Environmental Controls, and the Partitioning of N₂O Sources in UK Natural and Seminatural Land Use Types

Fotis Sgouridis¹ and Sami Ullah²
¹School of Geographical Sciences, University of Bristol, Bristol, UK, ²School of Geography, Earth and Environmental Sciences, and Birmingham Institute for Forest Research, University of Birmingham, Birmingham, UK

Abstract Natural and seminatural terrestrial ecosystems (unmanaged peatlands and forests and extensive and intensive grasslands) have been under-represented in the UK greenhouse gas (GHG) inventory. Mechanistic studies of GHG fluxes and their controls can improve the prediction of the currently uncertain GHG annual emission estimates. The source apportionment of N₂O emissions can further inform management plans for GHG mitigation. We have measured in situ GHG fluxes monthly in two replicated UK catchments and evaluated their environmental controlling factors. An adapted ¹⁵N-gas flux method with low addition of ¹⁵N tracer (0.03–0.5 kg ¹⁵N ha^{−1}) was used to quantify the relative contribution of denitrification to net N₂O production. Total N₂O fluxes were 40 times higher in the intensive grasslands than in the peatlands (range: −1.32 to 312.3 μg N m^{−2} h^{−1}). The contribution of denitrification to net N₂O emission varied across the land use types and ranged from 9 to 60%. Soil moisture was the key parameter regulating the partitioning of N₂O sources ($r^2 = 0.46$). Total N₂O fluxes were explained by a simple model ($r^2 = 0.83$) including parameters such as total dissolved nitrogen, organic carbon, and water content. A parsimonious model with the soil moisture content as a single scalar parameter explained 84% of methane flux variability across land uses. The assumption that 1% of the atmospherically deposited N on natural ecosystems is emitted as N₂O could be overestimated or underestimated (0.3–1.6%). The use of land use-specific N₂O emission factors and further information on N₂O source partitioning should help constrain this uncertainty.

1. Introduction

Natural and seminatural terrestrial ecosystems (peatlands, forests, and grasslands) can play a significant role as nonagricultural sources or sinks of the main greenhouse gases (GHG), carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O) (Dalal & Allen, 2008; Levy et al., 2012; Mander et al., 2010; Skiba et al., 2012). In the UK, 49% and 85% of the rural land cover in England and Wales, respectively, is composed of natural and seminatural land use types (Morton et al., 2011), which are currently collectively considered as net carbon sinks (land use, land use change, and forestry sector −9 Mt CO₂e in 2014; Brown et al., 2016). However, soil GHG emissions from nonfertilized native woodlands, mature poorly drained forests, and natural peatlands (not artificially drained or rewetted) are not currently reported to the UK GHG inventory (Brown et al., 2016). Indirect emissions due to atmospheric N deposition on natural ecosystems are not included in the national GHG inventory either, despite the uncertainty associated with the effect of increasing N deposition (Fowler et al., 2004, 2015) for N₂O emissions from natural forests (Skiba et al., 1998, 1999) and peatlands (Sgouridis & Ullah, 2015). For grasslands, the default Intergovernmental Panel on Climate Change (IPCC) Tier 1 emission factor approach (Intergovernmental Panel on Climate Change (IPCC), 2006), based on internationally agreed protocols using simple equations, is currently used, but this is not accurate enough to reflect regional variability driven by differences in land management intensity (e.g., intensive and extensive grasslands). To fill these gaps, there is an ongoing effort to develop and implement Tier 2 reporting systems that are country-specific taking into account climatic and management differences, for natural and seminatural ecosystems for a more accurate estimate of the annual GHG balance (Sgouridis & Ullah, 2015; Skiba et al., 2013). Further improvement and refinement of the Tier 2 reporting systems and the evolution of Tier 3 top-down modeling approaches can greatly benefit from linking landscape-scale flux measurements from natural and seminatural land use types with easily measurable environmental scalar parameters (Skiba et al., 2012, 2013).

©2017. The Authors.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Soil GHG fluxes, and particularly N_2O and CH_4 , are compounded by large spatiotemporal variability with spikes of biogeochemical activity creating “hot spots” and “hot moments” (McClain et al., 2003). This high variability emanates from the environmental heterogeneity of the biotic (e.g., microbial consortia and vegetation) and abiotic (e.g., substrate availability, soil aeration/ moisture, temperature, and pH) controls of the biogeochemical processes involved in the production or consumption of GHGs, and it is particularly difficult to simulate with process models that are applicable across natural ecosystem types (Dalal & Allen, 2008). Global methane emission controls include soil moisture or water table depth, soil carbon content, soil temperature and pH, and vegetation composition (Carter et al., 2012; Levy et al., 2012; Turetsky et al., 2014), but the relative importance of these controls varies depending on the spatial and temporal scales investigated (Dise, 1993; Levy et al., 2012) and also the ecosystem type. Soil temperature and moisture are also postulated as universal controls of respiratory CO_2 production across ecosystems (Gritsch, Zimmermann, & Zechmeister-Boltenstern, 2015).

Biological nitrous oxide production in soils is attributed to the aerobic processes of autotrophic and heterotrophic nitrification and nitrifier denitrification and the anaerobic processes of denitrification and dissimilatory nitrate reduction to ammonium (DNRA), with nitrification and denitrification being quantitatively the most important (Baggs, 2011; Butterbach-Bahl et al., 2013). Soil reactive N (labile organic bound N and inorganic N compounds) availability has been advocated as the major driver of N_2O emissions from soils (Butterbach-Bahl et al., 2013), while soil moisture and temperature provide additional controls by regulating the availability of oxygen and microbial enzymatic activity, respectively (Butterbach-Bahl et al., 2013; Schaufler et al., 2010). The interaction of these environmental controls, particularly due to the opposing conditions (aerobic versus anaerobic) for N_2O production, and the contribution of other distal controlling factors such as soil organic carbon quality, soil pH, and texture (Saggar et al., 2013; Sgouridis & Ullah, 2014) commonly lead to weak relationships between N_2O fluxes and environmental variables measured in the field (Carter et al., 2012; Luo et al., 2012; Skiba et al., 2013), further hampering our ability to predict N_2O fluxes at the landscape scale and in response to changing environmental conditions. Despite the wealth of studies that have investigated GHG fluxes and their controlling factors, the majority have focused on one or two land use types and only a few have considered all major GHGs across a range of natural and seminatural land use types in the laboratory (Schaufler et al., 2010), in meta-analysis (Mander et al., 2010), and under field conditions (Cz  bel et al., 2010). To evaluate whether natural and seminatural ecosystems ought to be included in IPCC reporting, particularly under a changing climate, further field measurements are needed. These in situ measurements need to follow a replicated catchment approach for deriving relatively more robust landscape-scale predictors both within and across ecosystem types in an attempt to reduce the uncertainties in regional-scale modeling.

Attributing N_2O emissions to different source pathways can further improve our understanding of the controls on the different processes and ultimately inform mitigation strategies given that N_2O is produced by many competing or coupled processes (Baggs, 2008, 2011). Soil moisture appears to be a key factor determining the partitioning of N_2O emissions between its sources (Baggs, Smales, & Bateman, 2010; Bateman & Baggs, 2005) and when denitrification will proceed to completion with the reduction of N_2O to N_2 (Davidson et al., 2000). Recent methodological developments, reviewed in Baggs (2008) and Butterbach-Bahl et al. (2013), have allowed N_2O source partitioning to be applied under field conditions. While natural abundance stable isotope approaches (P  rez et al., 2006) are noninvasive and can provide valuable information on which processes may be involved in N_2O production, they are inconclusive on their own and cannot provide quantification information (Baggs, 2008). Stable isotope enrichment approaches that involve the addition of $^{15}\text{N}\text{-NH}_4^+$ and/or $^{15}\text{N}\text{-NO}_3^-$ to soil have been the most reliable quantification method of N_2O source partitioning so far, but the need for large amounts of ^{15}N tracer application has restricted their use to fertilized agro-ecosystems (Baggs et al., 2003; Bateman & Baggs, 2005; Li et al., 2016; Mathieu et al., 2006) and forests receiving high rates of atmospheric N deposition (Ambus, Zechmeister-Boltenstern, & Butterbach-Bahl, 2006; Zhu et al., 2013). However, it was recently shown that low ^{15}N tracer application methods can be used in situ to quantify N_2 and N_2O fluxes from natural and seminatural ecosystems (Sgouridis, Stott, & Ullah, 2016; Sgouridis & Ullah, 2015) and this approach could be further expanded to account for N_2O source partitioning (Buchen et al., 2016; Morse & Bernhardt, 2013).

This paper reports in situ GHG fluxes from natural and seminatural ecosystems in two replicated UK catchments and elucidates the relative contribution of denitrification to net N_2O production. Our specific

objectives were to (1) assess the relative magnitude of GHG fluxes across a wide range of natural and semi-natural land use types, (2) evaluate the environmental controlling factors of GHG fluxes within different land use types, (3) attempt to derive simple models for predicting GHG fluxes across ecosystem types, and (4) partition the N_2O flux between denitrification and other possible sources and assess the environmental controls affecting the source apportionment.

2. Methods

2.1. Study Sites

Two river catchments in the UK, the Conwy (area 345 km²; North Wales, 52°59'82"N, 3°46'06"W) and the Ribble-Wyre (area 1145 km²; NW England, 53°59'99"N, 2°41'79"W), were selected for this study. In both of these catchments more than 90% of land cover consists of natural and seminatural rural land use types (Morton et al., 2011). These catchments were chosen as priority catchments by the UK Natural Environment Research Council (NERC) Macronutrient Cycles Programme (www.itls.org.uk) for investigating the magnitude and spatiotemporal variation of N, C, and P fluxes between soil, water, and air under a changing climate and perturbed C cycle.

In the Conwy catchment, four study sites (C-PB = peat bog; C-UG = unimproved grassland; C-IG = improved grassland; and C-MW = mixed woodland) were selected (Figure S1a in the supporting information). The headwater part of the Conwy River catchment lies at an average altitude of 440 m above sea level and has an average rainfall of 2200–2400 mm yr⁻¹. The C-PB and C-UG are under a light grazing regime, less than one sheep per hectare, while the C-IG, characterized by seasonally waterlogged cambic stagnogley soils, is intensively grazed perennially by both sheep and cattle, while fertilizer (range: 100–200 kg N ha⁻¹) and manure are applied twice per year during spring and summer months (E. Ritchie, personal communication). The C-MW (mature mixed wood forest and currently unmanaged) is characterized by typical brown podzolic soils that are shallow and well drained, while bare rock is locally visible and steep slopes are common.

In the Ribble-Wyre catchment, four study sites (R-UG = unimproved grassland; R-IG = improved grassland; R-HL = heathland; and R-DW = deciduous woodland) were selected (Figure S1b in the supporting information). The dominant soils in the area have been described as stagnopodzols to stagnohumic gleys, the altitude ranges from 260 to 290 m above sea level, and the average rainfall is 1693 mm yr⁻¹. The R-UG and R-IG have land management practices analogous to the ones described for the unimproved and improved grasslands in the Conwy catchment. Moreover, the R-UG was fertilized with N in the last decade once and has not been fertilized since, while it is being mowed twice per year (R. Rhodes, personal communication). The R-HL is privately owned, managed as a grouse moor, and grazed by sheep at low densities, while some recreational activities such as hiking are also allowed (Abbeystead Est., personal communication). The R-DW is an old growth forest developed on poorly drained soils and has never been fertilized (M. Colledge, personal communication). Further details on dominant plant species and soil texture of the study sites can be found in Sgouridis and Ullah (2014).

2.2. Gas Flux Measurements

Greenhouse gas fluxes were measured monthly using static chambers between April 2013 and October 2014 (total 17 months) with the exception of November 2013 and January 2014, which were not sampled. The measurement of greenhouse gas fluxes was part of a broader study aimed at quantifying in situ N_2 and N_2O fluxes due to denitrification from the same land use types (Sgouridis & Ullah, 2015) using an adapted ¹⁵N gas flux method (Mosier & Klemetsson, 1994) for low ¹⁵N tracer application, appropriate for natural and seminatural terrestrial ecosystems, which is described in detail in Sgouridis et al. (2016). Briefly, five plots were randomly established in each study site within each catchment amounting to 40 plots per monthly sampling campaign. In each plot a round PVC collar (basal area 0.05 m²; chamber volume 4 L) was inserted into the soil at ~10 cm depth (15 cm for the R-HL and C-PB plots) 2–4 weeks before the measurement date. The natural vegetation cover at the soil surface of each installed collar remained unchanged. The PVC collars were fitted with a circular groove of 25 mm depth to fit in an acrylic cylindrical cover (chamber) providing a gas-tight seal when filled with water, and the chamber was covered with reflective foil for minimizing temperature increase within the chamber headspace during the incubation period (Ullah & Moore, 2011). Labeled $\text{K}^{15}\text{NO}_3^-$ (98 at. % ¹⁵N, Sigma-Aldrich) was applied in each plot at a mean rate between 0.03 (±SE 0.005)

and $0.50 (\pm \text{SE } 0.073) \text{ kg N ha}^{-1}$ via 10 injections of equal volume through a grid ($4 \times 6 \text{ cm}$) using custom-made 10 cm long lumber needles (15 cm for the R-HL and C-PB plots) attached to a plastic syringe (Rutting, Boeckx, et al., 2011). The tracer solution (50–200 mL) was adjusted between 3 and 5% of the ambient volumetric water content. For the natural land use types, the average tracer application rate reflected the current daily estimates of atmospheric N deposition in the UK ($0.05 \text{ kg N ha}^{-1} \text{ d}^{-1}$) (Payne, 2014), while for the grassland soils the tracer application mimicked a daily fertilizer application rate of $0.5 \text{ kg N ha}^{-1} \text{ d}^{-1}$. The application of the ^{15}N tracer at these low rates should not be expected to enrich the soil nitrate pool significantly and potentially affect greenhouse gas emissions, in excess of the amount of nitrogen normally deposited via natural processes and common management practices in these land use types (Sgouridis et al., 2016). Following the ^{15}N tracer application, the collars were covered with the acrylic chamber fitted with a rubber septum for gas sampling. Two sets of gas samples (20 mL each) were collected with a gas tight syringe (SGE Analytical science) through the septum of the chamber cover at $T = 1 \text{ h}$ and $T = 2 \text{ h}$, while a $T = 0 \text{ h}$ sample was collected immediately after tracer injection above the plot surface before fitting the chamber cover. The gas samples were transferred into preevacuated ($<100 \text{ Pa}$) 12 mL borosilicate glass vials with butyl rubber septa (Exetainer vial; Labco Ltd., High Wycombe, United Kingdom) for storage under positive pressure and were analyzed within 8 weeks from collection without any significant change of the gas concentration (Laughlin & Stevens, 2003).

One set of the gas samples was analyzed for $^{15}\text{N-N}_2$ and $^{15}\text{N-N}_2\text{O}$ on an “in-house” laboratory-designed continuous flow isotope ratio mass spectrometer (CF-IRMS), and flux rates were calculated using the “non-equilibrium” equations as described in Sgouridis et al. (2016). The second set of gas samples was analyzed for total N_2O ($^{14} + ^{15}\text{N-N}_2\text{O}$) on a GC- μECD and for CH_4 and CO_2 on a GC-FID (7890A GC Agilent Technologies Ltd., Cheshire, UK). The minimum detectable concentration difference (MDCD) was defined for each gas with repeated manual analyses of reference standards ($n = 8$, $\text{N}_2\text{O} = 0.3 \text{ ppm}$, $\text{CH}_4 = 2 \text{ ppm}$, and $\text{CO}_2 = 200 \text{ ppm}$) and was calculated using the following equation (Matson, Pennock, & Bedard-Haughn, 2009):

$$\text{MDCD} = \mu_{\text{pair diff}} + (2\sigma_{\text{pair diff}}) \quad (1)$$

where μ is the mean difference of all possible unique pairs of reference standards ($n = 28$) and σ is the standard deviation between sample pairs. The MDCD values were 0.008, 0.036, and 3.6 ppm for N_2O , CH_4 , and CO_2 , respectively, while instrument precision was $<1\%$ relative standard deviation for all three gases. Only samples that were above or below (in case of consumption) the MDCD value for each gas were used for flux calculations, and if they were not, they were considered as zero fluxes. Flux rates were determined by linear regression of gas concentrations in the chamber headspace (adjusted for standard temperature and pressure) between 0 and 2 h, multiplied by the chamber volume and divided by the chamber area and time of incubation. The minimum detectable fluxes were $0.34 \mu\text{g N}_2\text{O-N m}^{-2} \text{ h}^{-1}$, $0.68 \mu\text{g CH}_4\text{-C m}^{-2} \text{ h}^{-1}$, and $67.14 \mu\text{g CO}_2\text{-C m}^{-2} \text{ h}^{-1}$.

For estimating the ratio of denitrification-derived N_2O over the total N_2O emission ($\text{DN}_2\text{O}/\text{TN}_2\text{O}$), the DN_2O flux rates calculated from the CF-IRMS samples (Sgouridis & Ullah, 2015) were used. Where necessary, TN_2O flux rates were calculated based on 20 h incubation to correspond to the respective time interval of DN_2O fluxes. Sgouridis et al. (2016) have shown that due to the high spatial variability between plots within each land use type, mean DN_2O fluxes were not statistically different when estimated based on 1, 2, or 20 h incubation. Therefore, an assumed N_2O flux linearity during 20 h of incubation did not significantly affect flux rate estimation due to the high spatial variability of N_2O emissions, which is in line with the findings by Chadwick et al. (2014). When TN_2O fluxes were negative (indicating consumption), but the proportion of the denitrification-derived N_2O (DN_2O) increased in the same sample, then all N_2O was assumed to be produced by denitrification. In case of zero TN_2O fluxes but detectable DN_2O flux, due to the lower limit of detection of the CF-IRMS ($0.2 \text{ ng N}_2\text{O-N m}^{-2} \text{ h}^{-1}$), then all N_2O was assumed to be produced by denitrification. In the occasions where DN_2O flux was larger than the TN_2O flux (10% of total sample pool), then 100% of the N_2O flux was assumed to derive from denitrification. Finally, in 3% of the samples, DN_2O was not measured successfully due to sampling or analytical bias, and in these cases the ratio of $\text{DN}_2\text{O}/\text{TN}_2\text{O}$ was not estimated.

Annual greenhouse gas fluxes were estimated by interpolating monthly measurements for each year and calculating the average over the two monitoring years. The global warming potential (GWP) for each land use type was calculated for the 100 year horizon, including climate-carbon feedback, as CO_2 equivalent by

multiplying average annual N_2O and CH_4 fluxes with the conversion factors of 298 and 34, respectively (IPCC, 2013).

Five composite soil samples (0–10 cm) were collected with a hand auger from each study site after the end of each gas flux measurement within 50 cm of each plot. The samples were transported to the laboratory on ice and stored at 4°C overnight. The analytical procedures for determining key soil properties are detailed in Text S1 in the supporting information.

2.3. Statistical Analysis

Factor analysis was used in Sgouridis and Ullah (2015) to cluster individual sites into distinct land use types. The sites C-PB, C-UG, and R-HL formed a distinct group called organic soils (OS), the sites C-IG and R-IG clustered together, forming the improved grassland (IG) land use type, while the site R-UG formed a separate land use type, named as semi-improved grassland (SIG). Finally, the mixed woodland (MW) in the Conwy catchment and the deciduous woodland (DW) in the Ribble-Wyre catchment were also considered as separate natural land use types. All subsequent results and statistical tests refer to the above land use type groups. Three-way analysis of variance (ANOVA) was performed for comparing the variance of the means between land use types, seasons, and years. Multiple linear regression was used for exploring the factors controlling greenhouse gas fluxes within and across land use types. All statistical analyses were performed using SPSS® 23.0 for Windows (IBM Corp., 2015, Armonk, NY) and are described in Text S2 in the supporting information.

3. Results

3.1. Soil Properties

The organic soil (OS) land use type (peat-bog, acid grassland, and heathland) was characterized by high soil organic matter and moisture contents and low bulk density, soil nitrate, and pH (Table 1). Conversely, the improved grasslands (IG) had significantly higher bulk density, nitrate, and ammonium contents as well as labile carbon availability (as indicated by the higher CO_2 production rate). The mixed woodland (MW) in the Conwy catchment, even though unmanaged, had similar nitrate and ammonium contents to the improved grasslands, but significantly lower soil moisture and pH compared to the IG (Table 1). The site R-UG formed a distinct land use type, named as semi-improved grassland (SIG), with intermediate soil properties between OS and IG (Table 1). Finally, the deciduous woodland (DW) in the Ribble-Wyre catchment had significantly higher soil pH, while it displayed similar soil moisture and nitrate contents to the SIG.

Apart from the highly significant differences of the soil properties between the five land use types (Table S1 in the supporting information), certain soil properties displayed significant seasonal variation most notably the water-filled pore space (WFPS), soil nitrate content, and soil temperature shown in Table S2 in the supporting information. The seasonal variation pattern was similar between all land use types with decreasing WFPS from winter to summer and increasing soil nitrate in the same seasonal order, while the soil temperature displayed a typical temperate seasonal cycle. The three-way ANOVA indicated significant intra-annual variation for the soil moisture, nitrate, dissolved organic carbon (DOC), and total dissolved nitrogen (TDN) contents with the second year having lower moisture (drier soil conditions), nitrate and TDN, and higher DOC values (data not shown).

3.2. Nitrous Oxide Fluxes and the Ratio $\text{DN}_2\text{O}/\text{TN}_2\text{O}$

The mean total N_2O fluxes (from all possible sources) were significantly different among land use types (ANOVA; $F = 171.4$, d.f. = 4, $p < 0.001$) and ranged between -1.32 and $312.3 \mu\text{g N m}^{-2} \text{ h}^{-1}$ (Figure 1a). Consumption of nitrous oxide (significant negative fluxes) was exclusively observed in the OS land use type, but on average OS was a minor source of N_2O about 40 times smaller than the IG, which emitted 3 times more N_2O than the SIG. The mean contribution of denitrification to the total N_2O flux ($\text{DN}_2\text{O}/\text{TN}_2\text{O}$) ranged between 0.09 and 0.59 and was lowest in the well-drained MW forest and highest in the poorly drained DW forest, while in the OS, IG, and SIG land uses was on average 0.44 (Figure 1a). The ratio $\text{DN}_2\text{O}/\text{TN}_2\text{O}$ increased seasonally in the order spring = summer < autumn < winter (Figure 1c). There was no consistent seasonal pattern of total N_2O emissions across the different land use types (Figure 1b), while event-related emissions were observed in both the SIG and IG land uses (e.g., N_2O emissions peaked after fertilization

Table 1
Soil Physico-chemical Properties in the Five Land Use Types in the Conwy and Ribble-Wyre River Catchments

	OS - organic soils (n = 255)	MW - mixed woodland (n = 85)	DW - deciduous woodland (n = 85)	SIG - semi-improved grassland (n = 85)	IG - improved grassland (n = 170)
Bulk density (g cm^{-3})	0.09 (0.006) ^a	0.56 (0.015) ^b	0.38 (0.010) ^c	0.29 (0.009) ^d	0.59 (0.008) ^b
WFPS (%)	69 (0.5) ^a	39 (1.0) ^b	65 (0.4) ^c	67 (1.0) ^{ac}	60 (0.8) ^d
Moisture content (% on w/w)	86 (0.5) ^a	38 (0.5) ^b	60 (0.6) ^c	64 (1.0) ^d	42 (0.5) ^e
pH	4.2 (0.03) ^a	5.2 (0.08) ^b	7.0 (0.04) ^c	5.8 (0.03) ^d	6.0 (0.01) ^e
Soil temperature ($^{\circ}\text{C}$)	10 (0.2) ^a	10 (0.3) ^{ab}	11 (0.3) ^b	12 (0.4) ^{bc}	12 (0.3) ^c
Organic matter (%)	90 (0.9) ^a	12 (0.3) ^b	25 (0.5) ^c	51 (1.4) ^d	18 (0.4) ^e
DOC (g m^{-2})	6.0 (0.27) ^a	4.3 (0.46) ^b	4.5 (0.30) ^b	10.5 (0.67) ^c	4.3 (0.23) ^b
CO ₂ production ($\text{mg C m}^{-2} \text{ h}^{-1}$)	43.8 (1.99) ^a	36.6 (2.00) ^a	53.3 (3.45) ^b	75.9 (4.87) ^c	160.3 (5.96) ^d
C:N (g g^{-1})	28 (0.8) ^a	11 (0.6) ^b	12 (0.5) ^b	18 (0.5) ^c	13 (0.6) ^b
NO ₃ ⁻ -N (g m^{-2})	0.02 (0.002) ^a	0.43 (0.046) ^b	0.14 (0.010) ^c	0.17 (0.013) ^c	0.38 (0.020) ^b
NH ₄ ⁺ -N (g m^{-2})	0.11 (0.012) ^a	0.32 (0.030) ^b	0.05 (0.007) ^c	0.19 (0.075) ^a	0.42 (0.053) ^b
TDN (g m^{-2})	0.37 (0.018) ^a	0.74 (0.052) ^b	0.41 (0.030) ^c	0.80 (0.059) ^b	0.80 (0.034) ^b

Note. Data are mean \pm standard error (SE) in parenthesis. Same lower case letters indicate no significant differences ($p > 0.05$) between land use types according to three-way ANOVA (one-way ANOVA for C:N ratio) and the Hochberg's GT2 post hoc test. For C:N ratio, $n = 30$ for organic soils, $n = 10$ for mixed and deciduous woodland and semiimproved grassland, and $n = 20$ for improved grassland. WFPS, water-filled pore space; DOC, dissolved organic carbon; TDN; Total dissolved nitrogen.

and/or rainfall events). The drier second sampling year (2014) led to significantly lower N₂O emissions particularly in the SIG and IG, while the ratio DN₂O/TN₂O was lower only in DW (Table S3).

The multiple linear regression (MLR) analysis indicated that 37% of the variance in total N₂O fluxes could be explained by the availability of labile organic carbon (as expressed by the soil respiration rate) and nitrate, with an additional 6% explained by soil temperature and pH amounting to 43% (Table 2). Averaging fluxes per sampling plot ($n = 40$), thus removing the temporal but retaining the spatial variability, resulted in a simpler regression model with significantly increased predictive power ($\log_{10}(\text{TN}_2\text{O} + 0.03) = 0.1 + 0.001 \times \text{TDN} + 0.007 \times \text{CO}_2 - 0.018 \times \text{volumetric water content (VWC)}$; $r^2 = 0.85$). The organic carbon availability was the key controlling factor of nitrous oxide emissions from the nitrate-rich soils of IG and MW, as well as the poorly drained DW forest soils. In the case of the organic carbon-rich soils of OS and SIG, higher N₂O emissions were associated with lower soil moisture and higher nitrate availability. The MLR analysis also showed that the ratio DN₂O/TN₂O is primarily controlled by the soil water content consistently across the different land use types, while secondary control factors include the soil pH, total dissolved nitrogen, and carbon availability (Table 2). In the poorly drained DW forest, showing the highest contribution of denitrification to N₂O emissions, the above soil parameters could explain 40% of the variance in the ratio DN₂O/TN₂O. Higher soil moisture content and pH also contributed to higher DN₂O/TN₂O ratio in the MW well-drained forest soils (explanatory power 26%). Aggregating fluxes per sampling plot increased the predictive power of soil moisture for the average ratio of DN₂O/TN₂O to 46%.

3.3. Methane Fluxes

The mean methane fluxes (Figure 2a) were significantly different between all the land use types (ANOVA; $F = 205.9$, d.f. = 4, $p < 0.001$) and ranged between -41.1 and $3474.5 \mu\text{g CH}_4\text{-C m}^{-2} \text{ h}^{-1}$. The OS, as expected, were net sources of CH₄ throughout the year (Figure 2b) with highest fluxes observed during summer and autumn months. The poorly drained soils of DW in the Ribble-Wyre catchment were also a significant CH₄ source particularly during winter. While the IGs were minor sinks of atmospheric CH₄, the SIG (representative of extensive grassland management) was a minor net source throughout the year (Figure 2b). Finally, the well-drained soils of MW in the Conwy catchment were net sinks of atmospheric CH₄. There was no significant difference of methane fluxes between the two monitoring years for most land use types, with the exception of DW and SIG which showed lower CH₄ emissions in 2014 (Table S3).

The MLR analysis of monthly methane fluxes across all land use types ($n = 680$) with selected soil physico-chemical variables (Table 2) showed that soil water content (measured as volumetric water content at 10 cm depth) can account for 59% of the variability in methane fluxes across natural and seminatural land use types, with an additional 3% explained by soil organic matter content and pH. MLR within each land

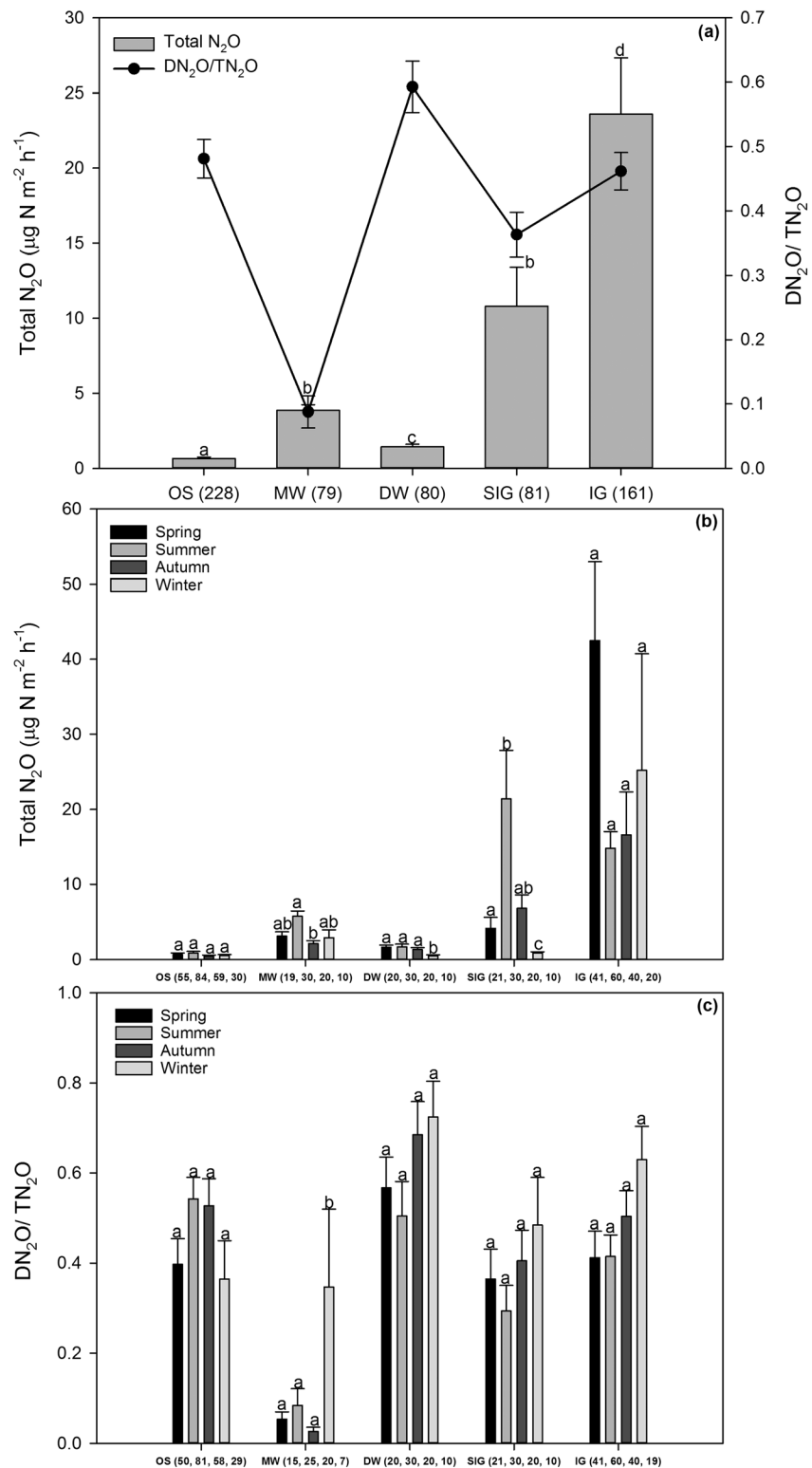


Figure 1. (a) Average total N₂O and the ratio DN₂O/TN₂O, (b) seasonal averages of total N₂O, and (c) seasonal averages of DN₂O/TN₂O in the five land use types in the Conwy and Ribble-Wyre River catchments. Same lower case letters indicate no significant differences ($p > 0.05$) between land use types (a) or seasons within land use types (b and c) according to one-way ANOVA and the Games-Howell post hoc test. The sample size (n) is given in parenthesis for each land use type on the x axis in the order (spring, summer, autumn, and winter). The error bars indicate $\pm 1SE$.

Table 2

Multiple Linear Regression Analysis Results Between (a) Methane Fluxes, (b) Carbon Dioxide Fluxes, (c) Total N₂O Fluxes, and (d) the Ratio DN₂O/TN₂O and Selected Soil Physico-chemical Properties

Dependent variable	Equation	r^2	n
(a) Methane			
All land use types	$-1084.5 + (13.8 \times \text{VWC}) + (3.5 \times \text{OM}) + (55.2 \times \text{pH})$	0.62	680
Organic soils	$-1311.2 + (146.8 \times \text{pH}) + (13.2 \times \text{VWC}) + (20.8 \times \text{T})$	0.40	255
Deciduous woodland	$169.8 - (65.6 \times \text{T}) + (150.4 \times \text{pH})$	0.60	85
Semi-improved grassland	$-1615 + (18.2 \times \text{VWC}) + (124.1 \times \text{pH})$	0.47	85
Improved grassland	$-40.4 - (25.2 \times \text{T}) + (4.9 \times \text{WFPS})$	0.39	170
(b) Carbon dioxide			
All land use types	$-180.6 + (13.3 \times \text{T}) + (111.9 \times \text{BD}) + (1.2 \times \text{WFPS})$	0.60	680
Organic soils	$-98.2 + (11.5 \times \text{T}) + (17 \times \text{pH}) - (0.7 \times \text{WFPS})$	0.69	255
Mixed woodland	$-164.9 + (11.9 \times \text{T}) + (1.8 \times \text{WFPS})$	0.58	85
Deciduous woodland	$-47.2 + (10.3 \times \text{T})$	0.49	85
Semi-improved grassland	$170.7 + (7.4 \times \text{T}) - (28.7 \times \text{pH})$	0.40	85
Improved grassland	$-94.8 + (13.1 \times \text{T}) + (3.8 \times \text{M})$	0.57	170
(c) Total N ₂ O			
All land use types	$-25.7 + (0.17 \times \text{CO}_2) + (0.03 \times \text{NO}_3^-) + (5.1 \times \text{pH}) - (1.2 \times \text{T})$	0.43	629
Organic soils	$57 - (0.45 \times \text{M}) - (8.2 \times \text{pH}) + (0.1 \times \text{NO}_3^-)$	0.20	228
Mixed woodland	$-21.8 + (0.41 \times \text{CO}_2) + (2 \times \text{T})$	0.46	79
Deciduous woodland	$-6.7 + (0.18 \times \text{CO}_2)$	0.21	80
Semi-improved grassland	$80.5 - (0.72 \times \text{WFPS}) + (0.07 \times \text{NO}_3^-) - (0.01 \times \text{TDN}) - (0.28 \times \text{OM})$	0.42	81
Improved grassland	$-123.3 + (0.13 \times \text{CO}_2) - (1.9 \times \text{T}) + (26.6 \times \text{pH})$	0.23	161
(d) DN ₂ O/ TN ₂ O			
All land use types	$-0.88 + (0.02 \times \text{WFPS}) + (0.16 \times \text{pH}) + (0.001 \times \text{TDN}) + (0.002 \times \text{CO}_2)$	0.22	606
Mixed woodland	$-2.4 + (0.35 \times \text{pH}) + (0.03 \times \text{VWC})$	0.26	67
Deciduous woodland	$-1.03 + (0.05 \times \text{VWC}) - (0.004 \times \text{CO}_2) + (0.001 \times \text{TDN})$	0.40	80
Semi-improved grassland	$-0.55 + (0.04 \times \text{VWC})$	0.23	81
Improved grassland	$1.007 + (0.001 \times \text{NO}_3^-) + (0.001 \times \text{NH}_4^+) + (0.01 \times \text{VWC})$	0.17	160

Note. The GHG flux data sets (a, b, and c) were normalized using the two-step approach, while the ratio DN₂O/TN₂O (d) was log₁₀-transformed before the regression (for details see Text S2 in the supporting information). All regressions are significant at $p < 0.01$. VWC, volumetric water content; OM, organic matter content; T, soil temperature; WFPS, water-filled pore space; BD, bulk density; M, soil moisture; NO₃⁻, nitrate content; CO₂, soil respiration; TDN, total dissolved nitrogen content; NH₄⁺, ammonia content.

use type revealed some further land use type-specific relationships (Table 2). Soil pH appeared to be an important controlling factor of methane fluxes in organic soils (21% explanatory power), with increased emissions associated with higher soil pH, while soil water content and temperature combined explained an additional 19% of methane flux variability in OS. Soil temperature alone explained 57 and 36% of the methane flux variability in DW and IG respectively, by indirectly affecting soil water availability during summer months (Figure 2b). Finally, in SIG the soil water content and pH combined accounted for 47% of the variability in methane fluxes. When the monthly fluxes were averaged per plot ($n = 40$) the soil water content alone explained 84% of the methane flux variation ($\log_{10}(\text{CH}_4 + 20) = -0.936 + 0.05 \times \text{VWC}$; $r^2 = 0.84$).

3.4. Carbon Dioxide Fluxes

The rate of CO₂ production (including soil microbial respiration, roots, and that of any aboveground vegetation enclosed by the static chamber) differed significantly between natural and seminatural land use types (ANOVA; $F = 117.8$, d.f. = 4, $p < 0.001$) and ranged between 0.63 and 435.5 mg C m⁻² h⁻¹. The IG emitted 4 times more carbon dioxide than the OS and MW land uses and twice as much as the SIG (extensively managed) (Figure S2a in the supporting information). The seasonal variability was consistent across all the land use types with larger CO₂ emissions observed in the summer and lowest in winter months (Figure S2b in the supporting information), while there were slightly higher emissions in the second drier year. Consequently, the soil temperature was shown to be the main controlling factor of CO₂ emissions across land uses ($\log_{10}(\text{CO}_2) = -1.053 + 0.26 \times \text{T}$; $r^2 = 0.86$). The soil temperature was consistently the most important controlling factor highlighted by MLR analysis within each land use type (Table 2), while the soil moisture was also a significant regulator in the drier IG and MW sites.

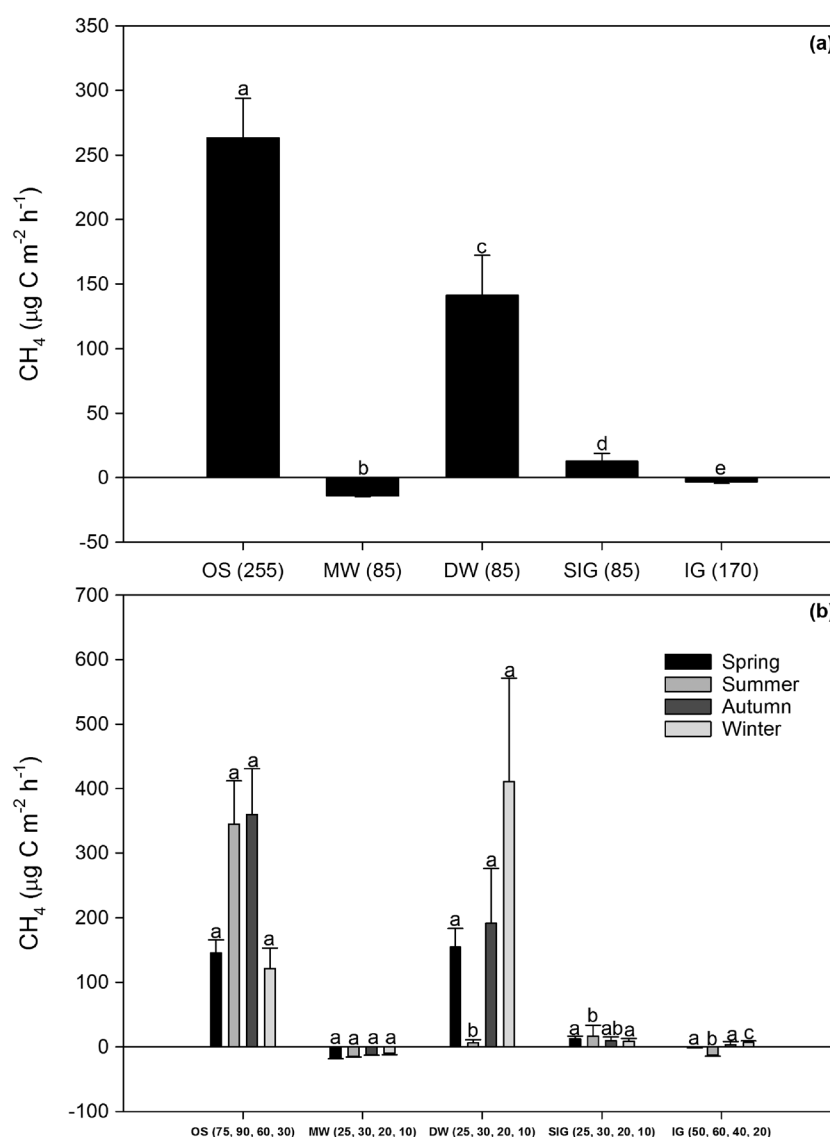


Figure 2. Methane fluxes: (a) mean rates and (b) seasonal averages in the five land use types in the Conwy and Ribble-Wyre River catchments. Same lower case letters indicate no significant differences ($p > 0.05$) between land use types (a) or seasons within land use types (b) according to one-way ANOVA and the Games-Howell *post hoc* test. The sample size (n) is given in parenthesis for each land use type on the x axis in the order (spring, summer, autumn, and winter). The error bars indicate $\pm 1\text{SE}$.

3.5. Annual Fluxes and Global Warming Potential

The average annual CH_4 flux across the natural and seminatural land use types ranged between -0.9 and $18.7 \text{ kg C ha}^{-1} \text{ yr}^{-1}$, while the respective annual N_2O flux range was 0.05 – $1.98 \text{ kg N ha}^{-1} \text{ yr}^{-1}$. The greenhouse gas budget for each land use type, calculated using the 100 year horizon global warming potential (GWP), was dominated by the soil and ground vegetation respiration of carbon dioxide (Table 3), while the net ecosystem exchange of CO_2 was not measured in this study. The contribution of both methane and nitrous oxide to the GWP was highest in the IG, almost entirely due to N_2O emission, followed by the OS, where CH_4 emission contributed 97.5% of the GWP. Methane was also the main contributor (94%) to GWP from the poorly drained forest soils of DW, whereas methane consumption in the well-drained forest soils of MW reduced the GWP by 49%. In the extensively managed grassland (SIG) both greenhouse gases contributed considerably to the GWP, which was almost 3 times lower than the GWP of the improved grasslands.

Table 3

Annual Average Greenhouse Gas Fluxes, Total Global Warming Potential (GWP_{total}), and the Contribution of Methane and Nitrous Oxide to the Global Warming Potential ($GWP_{N_2O + CH_4}$) in the Five Land Use Types in the Conwy and Ribble-Wyre River Catchments

Land use types	Greenhouse gas fluxes ($kg\ CO_{2eq}\ ha^{-1}\ yr^{-1}$)					$N_2O\ (kg\ N\ ha^{-1}\ yr^{-1})$	$CH_4\ (kg\ C\ ha^{-1}\ yr^{-1})$
	N_2O	CH_4	CO_2	GWP_{total}	$GWP_{N_2O + CH_4}$		
Organic soils	22.1	849.8	11044.2	11916	872	0.05	18.7
Mixed woodland	123.5	−40.6	9419.4	9502	83	0.26	−0.9
Deciduous woodland	48.2	733.1	13677.6	14459	781	0.10	16.2
Semi-improved grassland	302.2	51.6	18988.7	19343	354	0.65	1.1
Improved grassland	928.9	−0.2	39531.0	40460	929	1.98	0.0

4. Discussion

4.1. Nitrous Oxide Fluxes and the Ratio DN_2O/TN_2O

The application of ^{15}N tracer for measuring in situ the flux of N_2 and N_2O due to denitrification (Sgouridis et al., 2016; Sgouridis & Ullah, 2015) also enabled the estimation of the ratio of denitrification-derived N_2O over the total N_2O emission (DN_2O/TN_2O) under field conditions and for the first time across a wide spatial and temporal range in natural terrestrial ecosystems. The ^{15}N - N_2O emitted from a uniformly labeled soil NO_3^- pool can be attributed to the process of denitrification (DN_2O) when assuming negligible dissimilatory nitrate reduction to ammonium (DNRA) and immobilization and remineralization of ^{15}N - NO_3^- (Buchen et al., 2016; Zhu et al., 2013). Currently, there is no available methodology to discriminate N_2O emissions between denitrification and DNRA (Baggs, 2008, 2011), and for this study we have assumed negligible contribution from DNRA, which when quantitatively important in temperate ecosystems can constitute up to 10% of the denitrification activity (Rutting, Huygens, et al., 2011; Sgouridis et al., 2011). The unlabeled N_2O flux could be due to the contributions from autotrophic or heterotrophic nitrification and/or nitrifier denitrification (Buchen et al., 2016; Zhu et al., 2013), and further discrimination between these sources would have required double tracer labeling (Baggs et al., 2003; Bateman & Baggs, 2005; Mathieu et al., 2006; Morse & Bernhardt, 2013) and/or the use of natural abundance isotopomer approaches (Mander et al., 2014; Schmidt et al., 2004; Sutka et al., 2006) which, although desirable, was beyond the scope of this study.

The highest (mean 0.59) contribution of denitrification to the emission of N_2O was observed in the poorly drained DW forest, while the lowest (mean 0.09) was measured in the well-drained MW forest (Figure 1a). Soil moisture has been shown as the key factor regulating the partitioning of N_2O between denitrification and nitrification sources (Baggs et al., 2010; Bateman & Baggs, 2005; Li et al., 2016; Mathieu et al., 2006; Morse & Bernhardt, 2013), and this was confirmed by the MLR analysis in all the land use types (Table 2). In the case of the MW forest, which displayed the lowest WFPS and the highest availability of mineral N (Table 1), there was strong indication that nitrification rather than denitrification may be quantitatively more important as an N_2O source (Ambus et al., 2006; Matson et al., 2009; Ullah & Moore, 2011). This is also supported by the highest nitrification potential of the MW shown in a preliminary study (Sgouridis & Ullah, 2014). This suggests that enhanced nitrification due to increased N inputs in well-drained forest soils could consequently lead to increased N_2O emissions (Ambus et al., 2006). Conversely, the dominant N_2O source in the hydromorphic forest soils (mean WFPS 65%) of the DW was denitrification (Ullah & Moore, 2011), while nitrification activity may have also been limited by the lowest ammonium availability (Table 1). In the grasslands (IG and SIG), the average WFPS range between 60 and 67% led to higher N_2O emissions from nitrification sources (0.54–0.64), while the ratio of DN_2O/TN_2O increased with increasing soil moisture (Figure 1c and Table 2), which is in line with previous findings from laboratory incubation studies in grasslands (Bateman & Baggs, 2005; Mathieu et al., 2006; Stevens et al., 1997). Intermediate soil moistures (around 65% WFPS in grasslands) create optimal conditions for the simultaneous occurrence of nitrification and denitrification (Davidson et al., 2000) due to the uninhibited diffusion of both substrates and O_2 (Parton et al., 1996) and the occurrence of anaerobic microsites in soil aggregates (Sextstone et al., 1985). Our study highlights the quantitative importance of both nitrification and denitrification as N_2O sources under field conditions and also shows that their relative magnitude varies seasonally (Wolf & Brumme, 2002) driven by seasonal changes in soil moisture, which should be taken into account in mitigation strategies for reducing N_2O emissions from grasslands. Despite the high WFPS of the organic soils all year round (mean 69%), only 0.48 of the N_2O

emissions was attributed to denitrification with the ratio of $\text{DN}_2\text{O}/\text{TN}_2\text{O}$ rising to 0.54 and 0.52 in summer and autumn, respectively (Figure 1c). At 70% WFPS it has been shown that all of the N_2O emitted from grassland soils originated from denitrification (Bateman & Baggs, 2005), while at $\text{pH} < 4.5$ denitrification is quantitatively more important than nitrification (Baggs et al., 2010; Cheng et al., 2015). It is possible that nonhomogeneous distribution of the $^{15}\text{N}\text{-NO}_3^-$ tracer and potential gas loss through subsoil gas diffusion or further reduction of $^{15}\text{N}\text{-N}_2\text{O}$ to $^{15}\text{N}\text{-N}_2$ (Sgouridis et al., 2016) in the saturated highly porous peat soils may have led to underestimated contribution of denitrification to N_2O emissions from the OS. There is currently a lack of studies on source partitioning of N_2O emissions from natural peat/organic soils, with the exception of Pihlatie et al. (2004) where the unreliable acetylene inhibition technique (Baggs, 2008) was used for source partitioning. Our study is the first one to indicate that sources other than denitrification may contribute to N_2O emissions from this under-represented soil type. However, further investigation is warranted to elucidate the various potential pathways of N_2O emission using multiple tracer and/or natural abundance approaches.

Total nitrous oxide fluxes were highest from the improved grasslands, which emitted 3 times as much N_2O as the SIG extensively managed grassland (Imer et al., 2013; Skiba et al., 2013). The mean annual nitrous oxide flux rate from the improved grasslands ($2 \text{ kg N}_2\text{O-N ha}^{-1}$) corresponds to the lower range of the fluxes reported for similarly managed grasslands (N fertilization range: $150\text{--}200 \text{ kg N ha}^{-1} \text{ yr}^{-1}$) in the UK and Ireland (Cardenas et al., 2010; Rafique, Hennessy, & Kiely, 2011; Renou-Wilson et al., 2014; Skiba et al., 2013). The N_2O fluxes from the mixed and deciduous forest soils in this study ($0.1\text{--}0.3 \text{ kg N}_2\text{O-N ha}^{-1} \text{ yr}^{-1}$) were comparable to nitrous oxide fluxes from temperate and boreal forests (deciduous and coniferous) in Canada (Matson et al., 2009; Peichl et al., 2010; Ullah & Moore, 2011) under moderate atmospheric N deposition ($<20 \text{ kg N ha}^{-1} \text{ yr}^{-1}$), while they corresponded to the lower end of the fluxes reported for coniferous and deciduous European forests (Eickenscheidt et al., 2014; Luo et al., 2012; Pilegaard et al., 2006) at higher atmospheric N deposition ($20\text{--}35 \text{ kg N ha}^{-1} \text{ yr}^{-1}$). The mean annual N_2O efflux from the organic soils ($0.05 \text{ kg N}_2\text{O-N ha}^{-1} \text{ yr}^{-1}$) was significantly lower than from any other land use type and comparable only to effluxes from natural, pristine European peatlands (Carter et al., 2012; Leppelt et al., 2014), while it was 1–2 orders of magnitude lower than nitrous oxide emissions from organic soils used for agriculture (Petersen et al., 2012) or from drained peatlands (Mander et al., 2010) or peat extraction sites (Leppelt et al., 2014).

The multiple linear regression analysis indicated significant and relatively complex environmental controls of nitrous oxide emission from natural terrestrial ecosystems (Table 2) including nitrate and organic carbon availability, soil pH, and temperature with overall lower predictive power (43%) compared to the models predicting methane and carbon dioxide effluxes from the same land use types. Weak relationships between N_2O fluxes and environmental variables measured in the field are commonly found in the literature (Carter et al., 2012; Eickenscheidt et al., 2014; Luo et al., 2012; Skiba et al., 2013). This has been attributed to the highly dynamic nature of nitrous oxide production and consumption processes and their enormous spatiotemporal variability (Baggs, 2011; Butterbach-Bahl et al., 2013), which is difficult to predict particularly at weekly or monthly timescales (Luo et al., 2012). Aggregating fluxes per sampling plot resulted in a simpler regression model that encompasses the key environmental drivers of N availability in the form of total dissolved nitrogen concentration in soil water (69% predictive power), carbon availability as expressed by the soil respiration rate (12% predictive power), and soil moisture as VWC (4% predictive power) and is applicable across different natural terrestrial ecosystem types.

Soil reactive N (labile organic bound N and inorganic N compounds) availability has been advocated as the major driver of N_2O emissions from soils (Butterbach-Bahl et al., 2013), and the highly significant correlation ($r^2 = 0.69$; Figure 3a) between total N_2O fluxes and TDN highlights that not only nitrate (the substrate of denitrification) but also ammonium (the substrate of nitrification) and organic N forms (supply of nitrate and ammonium through mineralization) exert significant control over the emission of N_2O from multiple microbial processes. Sgouridis and Ullah (2014, 2015) have already shown that a gradient of N and organic carbon availability across the same natural and seminatural land use types, as this is influenced by management practices (e.g., fertilization, grazing, and hay cutting) in grasslands and natural variability in forests and organic soil, are the most significant proximal controls of denitrification activity and the associated N_2O emissions. Fertilization and grazing with mineral nitrogen in the improved grasslands are the primary reasons for the higher N_2O emissions from this land use type, and this dose-response relationship has been well documented in the literature (Cardenas et al., 2010; Rafique et al., 2011; Skiba et al., 2013) and likely explains

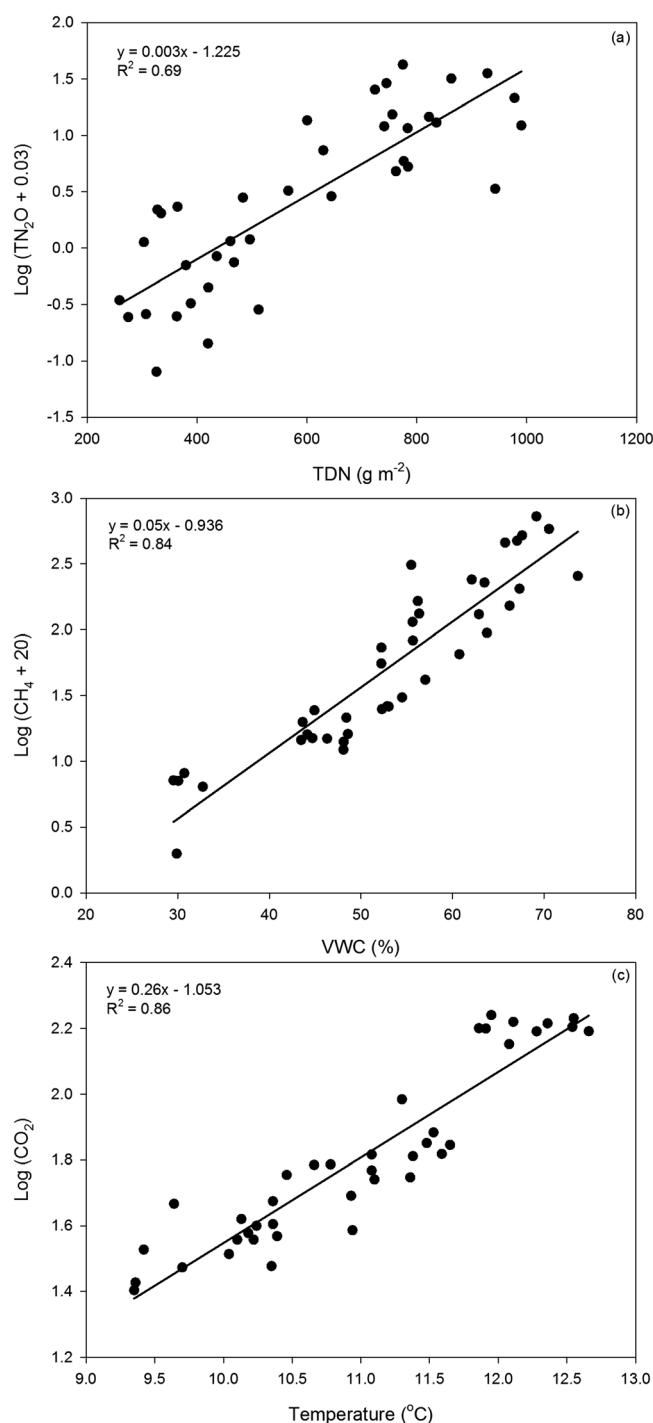


Figure 3. Correlations between (a) log₁₀-transformed total nitrous oxide fluxes and total dissolved nitrogen, (b) log₁₀-transformed methane fluxes and volumetric water content, and (c) log₁₀-transformed carbon dioxide fluxes and soil temperature per plot averaged over 17 months ($n = 40$). All correlations are significant at $p < 0.01$.

the 3 times lower emissions from the unfertilized SIG. Conversely, the N_2O emission from the OS was limited by nitrate supply as indicated by the MLR (Table 2) and also by the significantly lower nitrification and denitrification potentials compared to the other land use types (Sgouridis & Ullah, 2014).

4.2. Methane Fluxes

Land use type significantly affected methane fluxes, with the order of CH_4 emission rates being organic soils > poorly drained forest > semi-improved grassland > improved grassland > well-drained forest, which is in line with previous studies in rural ecosystems (Czóbel et al., 2010; Mander et al., 2010). The OSs (including an ombrotrophic peat bog, an acid grassland, and a heathland) were significant CH_4 sources at an average annual rate of $25 \text{ kg CH}_4 \text{ ha}^{-1}$, which corresponds to the lower range reported for organic soils in the UK (Levy et al., 2012), while it is lower than the mean flux from temperate pristine wetlands across the globe (Turetsky et al., 2014) and in Estonia (Mander et al., 2010). Poorly drained forest soils were the second most important CH_4 source (annual mean $21.6 \text{ kg CH}_4 \text{ ha}^{-1}$) at similar magnitude as deciduous forests on hydromorphic soils in the St. Lawrence River catchment, eastern Canada (Ullah & Moore, 2011), and in Estonia (Mander et al., 2010). Conversely, the well-drained soils of the mixed forest in the Conwy catchment were a small methane sink (annual mean $-1.2 \text{ kg CH}_4 \text{ ha}^{-1}$) comparable to methane uptake rates reported for automorphic forest soils in Europe (Luo et al., 2012; Mander et al., 2010) and Canada (Ullah & Moore, 2011). The improved grassland was on an annual basis methane neutral, while the semi-improved grassland was a small methane source (annual mean $1.5 \text{ kg CH}_4 \text{ ha}^{-1}$), results that generally agree with previous studies of the GHG balance of grasslands in similar ecoregions (Imer et al., 2013; Petersen et al., 2012; Renou-Wilson et al., 2014; Skiba et al., 2013).

The multiple linear regression analysis on averaged plots over time highlighted the soil water content as the key environmental driver (84% explanatory power) of methane flux across natural and seminatural land use types (Figure 3b). Previous studies have shown that averaging measurements over time can lead to stronger relationships between CH_4 flux and environmental variables as random measurement errors are canceled out (Levy et al., 2012), while temporal lags and “memory effects” in methanogenic activity are integrated (Turetsky et al., 2014). Soil water content is commonly used as a surrogate variable for soil O_2 concentration and is often a better proxy than water table depth for gauging the extent of methanogenesis (Imer et al., 2013; Levy et al., 2012). Significant methane emissions were measured at soil water contents between 55 and 70% VWC (in OS and DW sites), while methane consumption was more prominent at <40% VWC (MW site), which is in agreement with previous investigations of the relationship between methane emissions and soil moisture (Schaufli et al., 2010). Soil organic matter content explained an additional 2% of the variation in methane fluxes across

the land use types. Soil carbon availability has been shown to positively affect CH_4 emissions (Carter et al., 2012; Levy et al., 2012) directly through the supply of carbon substrate for methanogenesis, but also indirectly by increasing the water holding capacity of the soil (Hudson, 1994), thus leading to increased soil moisture content that hinders the aerobic decomposition of the available carbon (Jungkunst & Fiedler,

2007). A significant positive relationship was found between methane emission and soil pH in the OS, DW, and SIG sites (main methane sources among the investigated land use types), which for the organic soils in particular explained 21% of the methane flux variation. Increase in pH even by 0.5 (from 3.5 to 4.0) in peatlands and shrublands has been associated with increased CH_4 efflux by almost 300% (Carter et al., 2012), which was attributed to increased DOC availability due to the increased solubility of organic acids and humic substances in the soil brought about by the higher pH (Evans et al., 2008; Murakami, Furukawa, & Inubushi, 2005). Overall, our findings highlight the importance of soil water content as a broad spatial scale environmental driver of methane emissions from terrestrial ecosystems, which is easily measurable and can be used as a scalar indicator for predicting methane emissions at the catchment scale. Further testing of this simple model across land use types in several catchments is warranted for assessing its applicability for the development of Tier 3 emission factor models.

4.3. Carbon Dioxide Fluxes

The carbon dioxide fluxes from our static chamber measurements represent the combined flux from heterotrophic soil respiration and autotrophic respiration of the vegetation enclosed by the chamber, which constitute a significant proportion of the ecosystem exchange of CO_2 (Parkin et al., 2005). The CO_2 fluxes between land use types followed the order grasslands > forests > organic soils, and the mean annual range ($2.5\text{--}11\text{ t CO}_2\text{-C ha}^{-1}$) is within the ranges reported for similarly managed grasslands in the UK (Skiba et al., 2013), European peatlands and shrublands (Carter et al., 2012), and temperate forest soils in Canada (Ullah & Moore, 2011), while being lower than temperate forest soils in Europe (Luo et al., 2012). The addition of low amounts of ^{15}N tracer in the measured plots did not affect soil respiration rates as shown by Sgouridis et al. (2016), which is in agreement with previous studies on the effect of N fertilization on soil respiration (Carter et al., 2012; Liu & Greaver, 2009; Luo et al., 2012; Raich & Schlesinger, 1992; Skiba et al., 2013).

Soil temperature explained most of the variability (47%) of soil respiration across land use types, with an additional 13% explained by soil moisture and bulk density, which confirms the findings of several previous studies on the controls of soil CO_2 respiration in terrestrial ecosystems (Carter et al., 2012; Gritsch et al., 2015; Imer et al., 2013; Luo et al., 2012; Schauffer et al., 2010; Skiba et al., 2013). The temperature dependence of soil respiration can be explained by the stimulation of biological activity (plant roots and microbial communities) with increasing temperature, and this was evident within all the investigated land use types as indicated by the MLR analysis (Table 2). Temperature sensitivity is larger for relatively wet soils (WFPS: 60–70%) (Gritsch et al., 2015), which was shown for the OS and DW soils where soil temperature alone explained 67% and 49% of the CO_2 efflux variation. At very dry or very wet conditions soil moisture sensitivity over soil respiration also becomes important (Gritsch et al., 2015) and can limit CO_2 efflux either by restricting aeration and O_2 supply for aerobic respiration (wet conditions) or through osmotic stress of the microbial community under dry conditions (Smith et al., 2003). This could explain the observed negative relationship between soil respiration and WFPS in the OS sites (mean WFPS 70%) and the positive relationship between soil water content and CO_2 efflux in the MW and IG sites (mean WFPS 40–60%). When averaging measurements per sampling plot, soil temperature became the single scalar predictor (Figure 3c) of soil respiration across broad spatial scales within the soil water content range 40–70% WFPS, which is common across terrestrial ecosystems in northern latitudes.

4.4. Global Warming Potential and Emission Factors

The greenhouse gas (GHG) budget was estimated using the 100 year global warming potential, including climate-carbon feedback (IPCC, 2013), and for each land use type it was dominated by the CO_2 production (microbial plus root respiration, per se ecosystem respiration) representing more than 93% of the total GWP (Table 3). Of this ecosystem respiration term, 35–45% could be attributed to vegetation respiration (Silvola et al., 1996), particularly during the growth season between April and September, since plants were not removed from the collar enclosures. It has been shown that in similar land use types under comparable management regimes and climatic conditions (Petersen et al., 2012; Renou-Wilson et al., 2014; Skiba et al., 2013), the net ecosystem exchange of CO_2 is the dominant term of the GHG budget, with forests considered as annual net C sinks (Ullah & Moore, 2011), while in grasslands and organic soils the counteraction of the net ecosystem CO_2 exchange (NEE) by the emission of N_2O and CH_4 , respectively, varies spatiotemporally due to climatic and management effects. In this study we were not able to estimate the NEE or fluvial carbon

loss and biomass removal (Renou-Wilson et al., 2014) and therefore cannot conclusively argue whether the investigated land use types were net C sources or sinks. Moreover, manual monthly chamber-based measurements were interpolated to estimate annual GHG fluxes, and therefore, our GWP estimates are based on single-emission events propagated through time rather than continuous flux measurements. However, we were able to estimate the relative contribution of N_2O and CH_4 emission to GWP and have observed a similar GWP strength between organic soils and fertilized grasslands due almost entirely to methane and nitrous oxide emissions, respectively (Table 3). This finding highlights the need for climate change mitigation strategies not only for fertilized agro-ecosystems but also for natural and seminatural organic soils that, depending on climatic conditions, may be equal contributors to GWP, while their conversion for agricultural use may further lead to increased GWP contributions by both N_2O and CH_4 (Petersen et al., 2012). Poorly drained forest soils were the third highest GWP contributor (5% CH_4 and 0.3% N_2O), mainly due to methane emissions, and this should be taken into account in forest restoration strategies for C sequestration where an increased proportion of wetland forest soils has been suggested to optimize C sequestration (Ullah & Moore, 2011). Conversely, in well-drained forest soil, methane consumption was counteracted by a 33% contribution of N_2O emission to GWP reducing the source strength of GWP by an order of magnitude compared to the other land use types. Nitrous oxide emission factors calculated as a fraction of mineral and/or organic N inputs (fertilizer and manure, IG or manure application only, SIG) averaged at 0.4 and 0.9% for the semi-improved and improved grasslands respectively, which is at the lower end of the range of the default Tier 1 emission factor (0.3–3%; IPCC, 2006) and comparable to the emission factors estimated for fertilized seminatural grasslands in France (Klumpp et al., 2011).

Using simulated atmospheric N-deposition data for the UK (CEH Edinburgh, personal communication), we have estimated the fraction of N deposition-induced N_2O emissions from poorly drained forest (0.5%), well-drained forest (1.6%), and organic soils (0.3%) which corresponded to the lower range (0.2–5%) of the agriculture-related indirect N_2O emissions calculated by the IPCC Tier 1 methodology. The assumption that 1% of the deposited N on natural ecosystems is emitted as N_2O (IPCC, 2006) may overestimate this source in the case of organic soils or underestimate it in the case of N-rich well-drained forest soils, and the development of Tier 2 emission factors for natural and seminatural ecosystems, that take into account soil environmental conditions such as N and C availability and pH, is needed for more accurate prediction of N_2O emissions (Skiba et al., 2012).

5. Conclusion

The application of ^{15}N tracer allowed us to estimate the relative contribution of denitrification to the total N_2O emission under field conditions and for the first time across a wide spatial and temporal range in natural terrestrial ecosystems. Up to 60% of N_2O emission was attributed to denitrification, while the quantitative importance of both nitrification and denitrification as N_2O sources varied across land use types primarily driven by differences in soil moisture content. This finding can prove useful when designing mitigation strategies for the reduction of N_2O emission, particularly in grasslands, while further studies are needed to elucidate the quantitative importance of the various N_2O sources in soils under natural ecosystems (peatlands and forests) especially under increased atmospheric N deposition and drainage scenarios. A gradient of N and organic carbon availability across the natural and seminatural land use types, as this is influenced by management practices (e.g., fertilization, grazing, and hay cutting) in grasslands and natural variability in forests and organic soils, was the most significant proximal control of N_2O emission. The landscape heterogeneity of methane flux was primarily explained by the soil water content, while additional environmental controls such as organic carbon content, soil temperature, and pH can be used to constrain the variability within ecosystem types and should be considered when deriving Tier 2 and 3 methane emission factors. Methane emissions from organic soils and N_2O emissions from improved grasslands contributed equally to the GWP, indicating that not only managed but also natural and seminatural ecosystems should be considered in climate change mitigation strategies. For nitrous oxide emission factors in particular, the assumption that 1% of the atmospherically deposited N on natural ecosystems is emitted as N_2O may be overestimated in the case of natural organic soils or underestimated in the case of N-rich well-drained forest soils, and additional measurements in these natural land use types are needed to further constrain this uncertainty.

Acknowledgments

The authors are grateful to E. Ritchie and R. Rhodes for their collaboration and permission to access their land, as well as the National Trust in Conwy, the Abbeystead Estate in the Trough of Bowland, and the Forestry Commission in Gisburn Forest. We are also thankful to A. Dietrich, S. Dixon, and M. Reboul at Keele University for their help during field sampling and laboratory analysis; Andy Stott for his analytical assistance at the NERC's Stable Isotope Facility; and Ed Tipping (CEH Lancaster) for his guidance and suggestions for improving the manuscript. This research was funded by the UK Natural Environment Research Council (NERC) grant (NE/J011541/1) awarded to Keele University and supported by a "grant in kind" from the NERC Life Sciences Mass Spectrometry Facility Steering Committee. The authors declare no competing financial interest. Data are available through NERC's Environmental Information Data Centre (Ullah & Sgouridis, 2017; <https://doi.org/10.5285/d970c095-129a-41ac-9c82-950ab7804581>).

References

- Ambus, P., Zechmeister-Boltenstern, S., & Butterbach-Bahl, K. (2006). Sources of nitrous oxide emitted from European forest soils. *Biogeosciences*, 3(2), 135–145.
- Baggs, E. M. (2008). A review of stable isotope techniques for N₂O source partitioning in soils: Recent progress, remaining challenges and future considerations. *Rapid Communications in Mass Spectrometry*, 22(11), 1664–1672. <https://doi.org/10.1002/rcm.3456>
- Baggs, E. M. (2011). Soil microbial sources of nitrous oxide: Recent advances in knowledge, emerging challenges and future direction. *Current Opinion in Environmental Sustainability*, 3(5), 321–327. <https://doi.org/10.1016/j.cosust.2011.08.011>
- Baggs, E. M., Richter, M., Cadisch, G., & Hartwig, U. A. (2003). Denitrification in grass swards is increased under elevated atmospheric CO₂. *Soil Biology & Biochemistry*, 35(5), 729–732. [https://doi.org/10.1016/S0038-0717\(03\)00083-X](https://doi.org/10.1016/S0038-0717(03)00083-X)
- Baggs, E. M., Smales, C. L., & Bateman, E. J. (2010). Changing pH shifts the microbial sources as well as the magnitude of N₂O emission from soil. *Biology and Fertility of Soils*, 46(8), 793–805. <https://doi.org/10.1007/s00374-010-0484-6>
- Bateman, E. J., & Baggs, E. M. (2005). Contributions of nitrification and denitrification to N₂O emissions from soils at different water-filled pore space. *Biology and Fertility of Soils*, 41(6), 379–388.
- Brown, P., Broomfield, M., Buys, G., Cardenas, L., Kilroy, E., MacCarthy, J., ... Webb, N. (2016). UK Greenhouse Gas Inventory 1990 to 2014: Annual report for submission under the Framework Convention on Climate Change (Rep., 568 pp.). Oxfordshire, UK: Department of Energy and Climate Change.
- Buchen, C., Lewicka-Szczepak, D., Fuß, R., Helfrich, M., Flessa, H., & Well, R. (2016). Fluxes of N₂ and N₂O and contributing processes in summer after grassland renewal and grassland conversion to maize cropping on a Plaggic Anthrosol and a Histic Gleysol. *Soil Biology and Biochemistry*, 101, 6–19. <https://doi.org/10.1016/j.soilbio.2016.06.028>
- Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., & Zechmeister-Boltenstern, S. (2013). Nitrous oxide emissions from soils: How well do we understand the processes and their controls? *Philosophical Transactions of the Royal Society, B: Biological Sciences*, 368(1621). <https://doi.org/10.1098/rstb.2013.0122>
- Cardenas, L. M., Thorman, R., Ashlee, N., Butler, M., Chadwick, D., Chambers, B., ... Scholefield, D. (2010). Quantifying annual N₂O emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs. *Estimation of Nitrous Oxide Emission From Ecosystems and its Mitigation Technologies*, 136(3–4), 218–226. <https://doi.org/10.1016/j.agee.2009.12.006>
- Carter, M. S., Larsen, K. S., Emmett, B., Estiarte, M., Field, C., Leith, I. D., ... Beier, C. (2012). Synthesizing greenhouse gas fluxes across nine European peatlands and shrublands—Responses to climatic and environmental changes. *Biogeosciences*, 9(10), 3739–3755. <https://doi.org/10.5194/bg-9-3739-2012>
- Chadwick, D. R., Cardenas, L., Misselbrook, T. H., Smith, K. A., Rees, R. M., Watson, C. J., ... Dhanoa, M. S. (2014). Optimizing chamber methods for measuring nitrous oxide emissions from plot-based agricultural experiments. *European Journal of Soil Science*, 65(2), 295–307. <https://doi.org/10.1111/ejss.12117>
- Cheng, Y., Zhang, J.-B., Wang, J., Cai, Z.-C., & Wang, S.-Q. (2015). Soil pH is a good predictor of the dominating N₂O production processes under aerobic conditions. *Journal of Plant Nutrition and Soil Science*, 178(3), 370–373. <https://doi.org/10.1002/jpln.201400577>
- Czöbel, S., Horváth, L., Szirmai, O., Balogh, J., Pintér, K., Németh, Z., ... Tuba, Z. (2010). Comparison of N₂O and CH₄ fluxes from Pannonian natural ecosystems. *European Journal of Soil Science*, 61(5), 671–682. <https://doi.org/10.1111/j.1365-2389.2010.01275.x>
- Dalal, R. C., & Allen, D. E. (2008). Greenhouse gas fluxes from natural ecosystems. *Australian Journal of Botany*, 56(5), 369–407. <https://doi.org/10.1071/BT07128>
- Davidson, E. A., Keller, M., Erickson, H. E., Verchot, L. V., & Veldkamp, E. (2000). Testing a conceptual model of soil emissions of nitrous and nitric oxides. *Bioscience*, 50(8), 667–680.
- Dise, N. B. (1993). Methane emission from Minnesota peatlands: Spatial and seasonal variability. *Global Biogeochemical Cycles*, 7(1), 123–142. <https://doi.org/10.1029/92GB02299>
- Eickenscheidt, T., Heinichen, J., Augustin, J., Freibauer, A., & Drösler, M. (2014). Nitrogen mineralization and gaseous nitrogen losses from waterlogged and drained organic soils in a black alder (*Alnus glutinosa* (L.) Gaertn.) forest. *Biogeosciences*, 11(11), 2961–2976. <https://doi.org/10.5194/bg-11-2961-2014>
- Evans, C. D., Goodale, C. L., Caporn, S. J. M., Dise, N. B., Emmett, B. A., Fernandez, I. J., ... Sheppard, L. J. (2008). Does elevated nitrogen deposition or ecosystem recovery from acidification drive increased dissolved organic carbon loss from upland soil? A review of evidence from field nitrogen addition experiments. *Biogeochemistry*, 91(1), 13–35.
- Fowler, D., O'Donoghue, M., Muller, J. B., Smith, R. I., Dragosits, U., Skiba, U., ... Brimblecombe, P. (2004). A chronology of nitrogen deposition in the UK between 1900 and 2000. *Water, Air, & Soil Pollution: Focus*, 4(6), 9–23. <https://doi.org/10.1007/s11267-004-3009-1>
- Fowler, D., Steadman, C. E., Stevenson, D., Coyle, M., Rees, R. M., Skiba, U. M., ... Galloway, J. N. (2015). Effects of global change during the 21st century on the nitrogen cycle. *Atmospheric Chemistry and Physics*, 15(24), 13,849–13,893. <https://doi.org/10.5194/acp-15-13849-2015>
- Gardner, W. H. (1965). Water content. In C. A. Black, D. D. Evans, J. L. White, L. E. Ensminger, & F. E. Clark (Eds.), *Methods of Soil Analysis. Part 1: Physical and Mineralogical Properties, Including Statistics of Measurement and Sampling* (pp. 82–125). Madison, WI: American Society of Agronomy.
- Gritsch, C., Zimmermann, M., & Zechmeister-Boltenstern, S. (2015). Interdependencies between temperature and moisture sensitivities of CO₂ emissions in European land ecosystems. *Biogeosciences*, 12(20), 5981–5993. <https://doi.org/10.5194/bg-12-5981-2015>
- Heiri, O., Lotter, A. F., & Lemcke, G. (2001). Loss on ignition as a method for estimating organic and carbonate content in sediments: Reproducibility and comparability of results. *Journal of Paleolimnology*, 25, 101–110.
- Hudson, B. D. (1994). Soil organic matter and available water capacity. *Journal of Soil and Water Conservation*, 49(2), 189–194.
- Imer, D., Merbold, L., Eugster, W., & Buchmann, N. (2013). Temporal and spatial variations of soil CO₂, CH₄ and N₂O fluxes at three differently managed grasslands. *Biogeosciences*, 10(9), 5931–5945. <https://doi.org/10.5194/bg-10-5931-2013>
- IPCC (2006). IPCC guidelines for national greenhouse gas inventories, prepared by the national greenhouse gas inventories programme (Rep.), Hayama, Japan.
- IPCC (2013). Climate change 2013: The physical science basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change (Rep., 1535 pp.). Cambridge, UK and New York.
- Jungkunst, H. F., & Fiedler, S. (2007). Latitudinal differentiated water table control of carbon dioxide, methane and nitrous oxide fluxes from hydromorphic soils: Feedbacks to climate change. *Global Change Biology*, 13(12), 2668–2683. <https://doi.org/10.1111/j.1365-2486.2007.01459.x>
- Kirkwood, D. S. (1996). *Nutrients: Practical Notes on Their Determination in Seawater*. Copenhagen, Denmark: ICES.
- Klump, K., Bloor, J. M. G., Ambus, P., & Soussana, J.-F. (2011). Effects of clover density on N₂O emissions and plant-soil N transfers in a fertilised upland pasture. *Plant and Soil*, 343(1), 97–107. <https://doi.org/10.1007/s11104-010-0526-8>

- Laughlin, R. J., & Stevens, R. J. (2003). Changes in composition of nitrogen-15-labeled gases during storage in septum-capped vials. *Soil Science Society of America Journal*, 67(2), 540–543.
- Leppelt, T., Dechow, R., Gebbert, S., Freibauer, A., Lohila, A., Augustin, J., ... Strömberg, M. (2014). Nitrous oxide emission budgets and land-use-driven hotspots for organic soils in Europe. *Biogeosciences*, 11(23), 6595–6612. <https://doi.org/10.5194/bg-11-6595-2014>
- Levy, P. E., Burden, A., Cooper, M. D. A., Dinsmore, K. J., Drewer, J., ... Zielinski, P. (2012). Methane emissions from soils: Synthesis and analysis of a large UK data set. *Global Change Biology*, 18(5), 1657–1669. <https://doi.org/10.1111/j.1365-2486.2011.02616.x>
- Li, X., Sørensen, P., Olesen, J. E., & Petersen, S. O. (2016). Evidence for denitrification as main source of N₂O emission from residue-amended soil. *Soil Biology and Biochemistry*, 92, 153–160. <https://doi.org/10.1016/j.soilbio.2015.10.008>
- Linn, D. M., & Doran, J. W. (1984). Effect of water-filled pore space on carbon dioxide and nitrous oxide production in tilled and non-tilled soils. *Soil Science Society of America Journal*, 48, 1267–1272.
- Liu, L., & Greaver, T. L. (2009). A review of nitrogen enrichment effects on three biogenic GHGs: The CO₂ sink may be largely offset by stimulated N₂O and CH₄ emission. *Ecology Letters*, 12(10), 1103–1117. <https://doi.org/10.1111/j.1461-0248.2009.01351.x>
- Luo, G. J., Brüggemann, N., Wolf, B., Gasche, R., Grote, R., & Butterbach-Bahl, K. (2012). Decadal variability of soil CO₂, NO, N₂O, and CH₄ fluxes at the Högwald Forest, Germany. *Biogeosciences*, 9(5), 1741–1763. <https://doi.org/10.5194/bg-9-1741-2012>
- Mander, U., Uuemaa, E., Kull, A., Kanal, A., Maddison, M., Soosaar, K., ... Augustin, J. (2010). Assessment of methane and nitrous oxide fluxes in rural landscapes. *Landscape and Urban Planning*, 98(3–4), 172–181. <https://doi.org/10.1016/j.landurbplan.2010.08.021>
- Mander, U., Well, R., Weymann, D., Soosaar, K., Maddison, M., Kanal, A., ... Tournebise, J. (2014). Isotopologue ratios of N₂O and N₂ measurements underpin the importance of denitrification in differently N-loaded riparian alder forests. *Environmental Science & Technology*, 48(20), 11,910–11,918. <https://doi.org/10.1021/es501727h>
- Mathieu, O., Henault, C., Leveque, J., Baujard, E., Milloux, M. J., & Andreux, F. (2006). Quantifying the contribution of nitrification and denitrification to the nitrous oxide flux using ¹⁵N tracers. *Environmental Pollution*, 144(3), 933–940.
- Matson, A., Pennock, D., & Bedard-Haughn, A. (2009). Methane and nitrous oxide emissions from mature forest stands in the boreal forest, Saskatchewan, Canada. *Forest Ecology and Management*, 258(7), 1073–1083. <https://doi.org/10.1016/j.foreco.2009.05.034>
- McClain, M. E., Boyer, E. W., Dent, C. L., Gergel, S. E., Grimm, N. B., Groffman, P. M., ... Pinay, G. (2003). Biogeochemical hot spots and hot moments at the interface of terrestrial and aquatic ecosystems. *Ecosystems*, 6, 301–312.
- Morse, J. L., & Bernhardt, E. S. (2013). Using N-15 tracers to estimate N₂O and N-2 emissions from nitrification and denitrification in coastal plain wetlands under contrasting land-uses. *Soil Biology & Biochemistry*, 57, 635–643. <https://doi.org/10.1016/j.soilbio.2012.07.025>
- Morton, D., Rowland, C., Wood, C., Meek, L., Marston, C., Smith, G., ... Simpson, I. C. (2011). Final report for LCM2007—The new UK land cover Map (Rep.). Centre for Ecology & Hydrology.
- Mosier, A. R., & Klemetsson, L. (1994). Measuring denitrification in the field. In R. W. Weaver, J. S. Angle, & P. S. Bottomley (Eds.), *Methods of Soil Analysis. Part 2: Microbiological and Biochemical Properties* (pp. 1047). WI: Soil Science Society of America, Inc.
- Murakami, M., Furukawa, Y., & Inubushi, K. (2005). Methane production after liming to tropical acid peat soil. *Soil Science & Plant Nutrition*, 51(5), 697–699. <https://doi.org/10.1111/j.1747-0765.2005.tb00094.x>
- Parkin, T. B., Kaspar, T. C., Senwo, Z., Prueger, J. H., & Hatfield, J. L. (2005). Relationship of soil respiration to crop and landscape in the Walnut Creek watershed. *Journal of Hydrometeorology*, 6(6), 812–824. <https://doi.org/10.1175/JHM459.1>
- Parton, W. J., Mosier, A. R., Ojima, D. S., Valentine, D. W., Schimel, D. S., Weier, K., & Kulmala, A. E. (1996). Generalized model for N₂ and N₂O production from nitrification and denitrification. *Global Biogeochemical Cycles*, 10, 401–412.
- Payne, R. J. (2014). The exposure of British peatlands to nitrogen deposition, 1900–2030. *Mires and Peat*, 14, 04–04.
- Peichl, M., Arain, M. A., Ullah, S., & Moore, T. R. (2010). Carbon dioxide, methane, and nitrous oxide exchanges in an age-sequence of temperate pine forests. *Global Change Biology*, 16(8), 2198–2212. <https://doi.org/10.1111/j.1365-2486.2009.02066.x>
- Pérez, T., García-Montiel, D., Trumbore, S., Tyler, S., Camargo, P. D., Moreira, M., ... Cerri, C. (2006). Nitrous oxide nitrification and denitrification ¹⁵N enrichment factors from Amazon forest soils. *Ecological Applications*, 16(6), 2153–2167. [https://doi.org/10.1890/1051-0761\(2006\)016%2153:NONADN%2.0.CO;2](https://doi.org/10.1890/1051-0761(2006)016%2153:NONADN%2.0.CO;2)
- Petersen, S. O., Hoffmann, C. C., Schäfer, C. M., Blicher-Mathiesen, G., Elsgaard, L., Kristensen, K., ... Greve, M. H. (2012). Annual emissions of CH₄ and N₂O, and ecosystem respiration, from eight organic soils in Western Denmark managed by agriculture. *Biogeosciences*, 9(1), 403–422. <https://doi.org/10.5194/bg-9-403-2012>
- Pihlatie, M., Syväsalo, E., Simojoki, A., Esala, M., & Regina, K. (2004). Contribution of nitrification and denitrification to N₂O production in peat, clay and loamy sand soils under different soil moisture conditions. *Nutrient Cycling in Agroecosystems*, 70(2), 135–141. <https://doi.org/10.1023/B:FRES.0000048475.81211.3c>
- Pilegaard, K., Skiba, U., Ambus, P., Beier, C., Brüggemann, N., Butterbach-Bahl, K., ... Zechmeister-Boltenstern, S. (2006). Factors controlling regional differences in forest soil emission of nitrogen oxides (NO and N₂O). *Biogeosciences*, 3(4), 651–661.
- Rafique, R., Hennessy, D., & Kiely, G. (2011). Nitrous oxide emission from grazed grassland under different management systems. *Ecosystems*, 14(4), 563–582. <https://doi.org/10.1007/s10021-011-9434-x>
- Raich, J. W., & Schlesinger, W. H. (1992). The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus B*, 44(2), 81–99. <https://doi.org/10.1034/j.1600-0889.1992.t01.1-00001.x>
- Renou-Wilson, F., Barry, C., Müller, C., & Wilson, D. (2014). The impacts of drainage, nutrient status and management practice on the full carbon balance of grasslands on organic soils in a maritime temperate zone. *Biogeosciences*, 11(16), 4361–4379. <https://doi.org/10.5194/bg-11-4361-2014>
- Rowell, D. L. (1994). *Soil Science: Methods and Applications*. New York: Longman Scientific & Technical.
- Rutting, T., Boeckx, P., Mueller, C., & Klemetsson, L. (2011). Assessment of the importance of dissimilatory nitrate reduction to ammonium for the terrestrial nitrogen cycle. *Biogeosciences*, 8(7), 1779–1791. <https://doi.org/10.5194/bg-8-1779-2011>
- Rutting, T., Huygens, D., Staelens, J., Mueller, C., & Boeckx, P. (2011). Advances in N-15-tracing experiments: New labelling and data analysis approaches. *Biochemical Society Transactions*, 39, 279–283. <https://doi.org/10.1042/BST0390279>
- Saggar, S., Jha, N., Deslippe, J., Bolan, N. S., Luo, J., Giltrap, D. L., ... Tillman, R. W. (2013). Denitrification and N₂O:N₂ production in temperate grasslands: Processes, measurements, modelling and mitigating negative impacts. *Soil as a Source & Sink for Greenhouse Gases*, 465(0), 173–195. <https://doi.org/10.1016/j.scitotenv.2012.11.050>
- Schaufler, G., Kitzler, B., Schindlbacher, A., Skiba, U., Sutton, M. A., & Zechmeister-Boltenstern, S. (2010). Greenhouse gas emissions from European soils under different land use: Effects of soil moisture and temperature. *European Journal of Soil Science*, 61(5), 683–696. <https://doi.org/10.1111/j.1365-2389.2010.01277.x>
- Schmidt, H.-L., Werner, R. A., Yoshida, N., & Well, R. (2004). Is the isotopic composition of nitrous oxide an indicator for its origin from nitrification or denitrification? A theoretical approach from referred data and microbiological and enzyme kinetic aspects. *Rapid Communications in Mass Spectrometry*, 18(18), 2036–2040. <https://doi.org/10.1002/rcm.1586>

- Sextstone, A. J., Revsbech, N. P., Parkin, T. B., & Tiedje, J. M. (1985). Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal*, 49(3), 645–651.
- Sgouridis, F., & Ullah, S. (2014). Denitrification potential of organic, forest and grassland soils in the Ribble-Wyre and Conwy River catchments, UK. *Environmental Science: Processes and Impacts*, 16(7), 1551–1562. <https://doi.org/10.1039/c3em00693j>
- Sgouridis, F., & Ullah, S. (2015). Relative magnitude and controls of in situ N_2 and N_2O fluxes due to denitrification in natural and seminatural terrestrial ecosystems using ^{15}N tracers. *Environmental Science and Technology*, 49(24), 14,110–14,119. <https://doi.org/10.1021/acs.est.5b03513>
- Sgouridis, F., Heppell, C. M., Wharton, G., Lansdown, K., & Trimmer, M. (2011). Denitrification and dissimilatory nitrate reduction to ammonium (DNRA) in a temperate re-connected floodplain. *Water Research*, 45(16), 4909–4922. <https://doi.org/10.1016/j.watres.2011.06.037>
- Sgouridis, F., Stott, A., & Ullah, S. (2016). Application of the ^{15}N gas-flux method for measuring in situ N_2 and N_2O fluxes due to denitrification in natural and semi-natural terrestrial ecosystems and comparison with the acetylene inhibition technique. *Biogeosciences*, 13(6), 1821–1835. <https://doi.org/10.5194/bg-13-1821-2016>
- Silvola, J., Alm, J., Ahlholm, U., Nykänen, H., & Martikainen, P. J. (1996). The contribution of plant roots to CO_2 fluxes from organic soils. *Biology and Fertility of Soils*, 23(2), 126–131. <https://doi.org/10.1007/BF00336052>
- Skiba, U., Sheppard, L., Pitcairn, C. E. R., Leith, I., Crossley, A., Van Dijk, S., ... Fowler, D. (1998). Soil nitrous oxide and nitric oxide emissions as indicators of elevated atmospheric N deposition rates in seminatural ecosystems. *Environmental Pollution*, 102(SUPPL. 1), 457–461. [https://doi.org/10.1016/S0269-7491\(98\)80069-9](https://doi.org/10.1016/S0269-7491(98)80069-9)
- Skiba, U., Sheppard, L. J., Pitcairn, C. E. R., Van Dijk, S., & Rossall, M. J. (1999). The effect of N deposition on nitrous oxide and nitric oxide emissions from temperate forest soils. *Water, Air, and Soil Pollution*, 116(1–2), 89–98. <https://doi.org/10.1023/A:1005246625038>
- Skiba, U., Jones, S. K., Dragosits, U., Drewer, J., Fowler, D., Rees, R. M., ... Manning, A. J. (2012). UK emissions of the greenhouse gas nitrous oxide. *Philosophical Transactions of the Royal Society, B: Biological Sciences*, 367(1593), 1175–1185. <https://doi.org/10.1098/rstb.2011.0356>
- Skiba, U., Jones, S. K., Drewer, J., Helfter, C., Anderson, M., Dinsmore, K., ... Sutton, M. A. (2013). Comparison of soil greenhouse gas fluxes from extensive and intensive grazing in a temperate maritime climate. *Biogeosciences*, 10(2), 1231–1241. <https://doi.org/10.5194/bg-10-1231-2013>
- Smith, K. A., Ball, T., Conen, F., Dobbie, K. E., Massheder, J., & Rey, A. (2003). Exchange of greenhouse gases between soil and atmosphere: Interactions of soil physical factors and biological processes. *European Journal of Soil Science*, 54(4), 779–791. <https://doi.org/10.1046/j.1351-0754.2003.0567.x>
- Stevens, R. J., Laughlin, R. J., Burns, L. C., Arah, J. R. M., & Hood, R. C. (1997). Measuring the contributions of nitrification and denitrification to the flux of nitrous oxide from soil. *Soil Biology and Biochemistry*, 29(2), 139–151.
- Sutka, R. L., Ostrom, N. E., Ostrom, P. H., Breznak, J. A., Gandhi, H., Pitt, A. J., & Li, F. (2006). Distinguishing nitrous oxide production from nitrification and denitrification on the basis of isotopomer abundances. *Applied and Environmental Microbiology*, 72(1), 638–644. <https://doi.org/10.1128/AEM.72.1.638-644.2006>
- Templeton, G. F. (2011). A two-step approach for transforming continuous variables to normal: Implications and recommendations for IS research. *Communications of the Association for Information Systems*, 28(1), 41–58.
- Turetsky, M. R., Kotowska, A., Bubier, J., Dise, N. B., Crill, P., Hornibrook, E. R. C., ... Wilkening, M. (2014). A synthesis of methane emissions from 71 northern, temperate, and subtropical wetlands. *Global Change Biology*, 20(7), 2183–2197. <https://doi.org/10.1111/gcb.12580>
- Ullah, S., & Moore, T. R. (2011). Biogeochemical controls on methane, nitrous oxide, and carbon dioxide fluxes from deciduous forest soils in eastern Canada. *Journal of Geophysical Research*, 116, G03010. <https://doi.org/10.1029/2010JG001525>
- Ullah, S., & Sgouridis, F. (2017). Denitrification and greenhouse gas emissions in natural and semi-natural terrestrial ecosystems [LTLS], NERC Environmental Information Data Centre, <https://doi.org/10.5285/d970c095-129a-41ac-9c82-950ab7804581>
- Wolf, I., & Brumme, R. (2002). Contribution of nitrification and denitrification sources for seasonal N_2O emissions in an acid German forest soil. *Soil Biology and Biochemistry*, 34(5), 741–744. [https://doi.org/10.1016/S0038-0717\(02\)00001-9](https://doi.org/10.1016/S0038-0717(02)00001-9)
- Zhu, J., Mulder, J., Bakken, L., & Dörsch, P. (2013). The importance of denitrification for N_2O emissions from an N-saturated forest in SW China: Results from in situ ^{15}N labeling experiments. *Biogeochemistry*, 116(1), 103–117. <https://doi.org/10.1007/s10533-013-9883-8>